

### Introduction

- Colistin and polymyxin B (Polymyxins class) are used as last resort drugs to treat multidrug resistance (MDR) Gram-negative infections (1)
- Resistance to polymyxins are increasing and plasmid-carried resistance (MCR-1) in natural colistin susceptible Gram-negative bacteria (GNB) has been recently reported (2,3)
- In *E. coli* study, *mcr-1* mediated low-level of polymyxins resistance by encoded the expression of phosphoethanolamine in lipid A of bacterial cell membrane (4). Whether *mcr-1* could increase GNB resistance level has not been studied yet

### Purpose

- To decipher heterogeneity of bacterial population (existing resistant subpopulation in total population) or adaptation to polymyxins resistance
- To describe, by an original approach of sequential time-kill experiments, the role of MCR-1 in the development of additional adaptive resistance to polymyxins in MDR Gram-negative bacteria.

### Methods

- Escherichia coli* J53, *Klebsiella pneumoniae* R2292 and their transconjugant *mcr-1* (J53\_ *mcr1* & KP\_ *mcr1*) were used in this study which were kindly provided by P. Nordmann

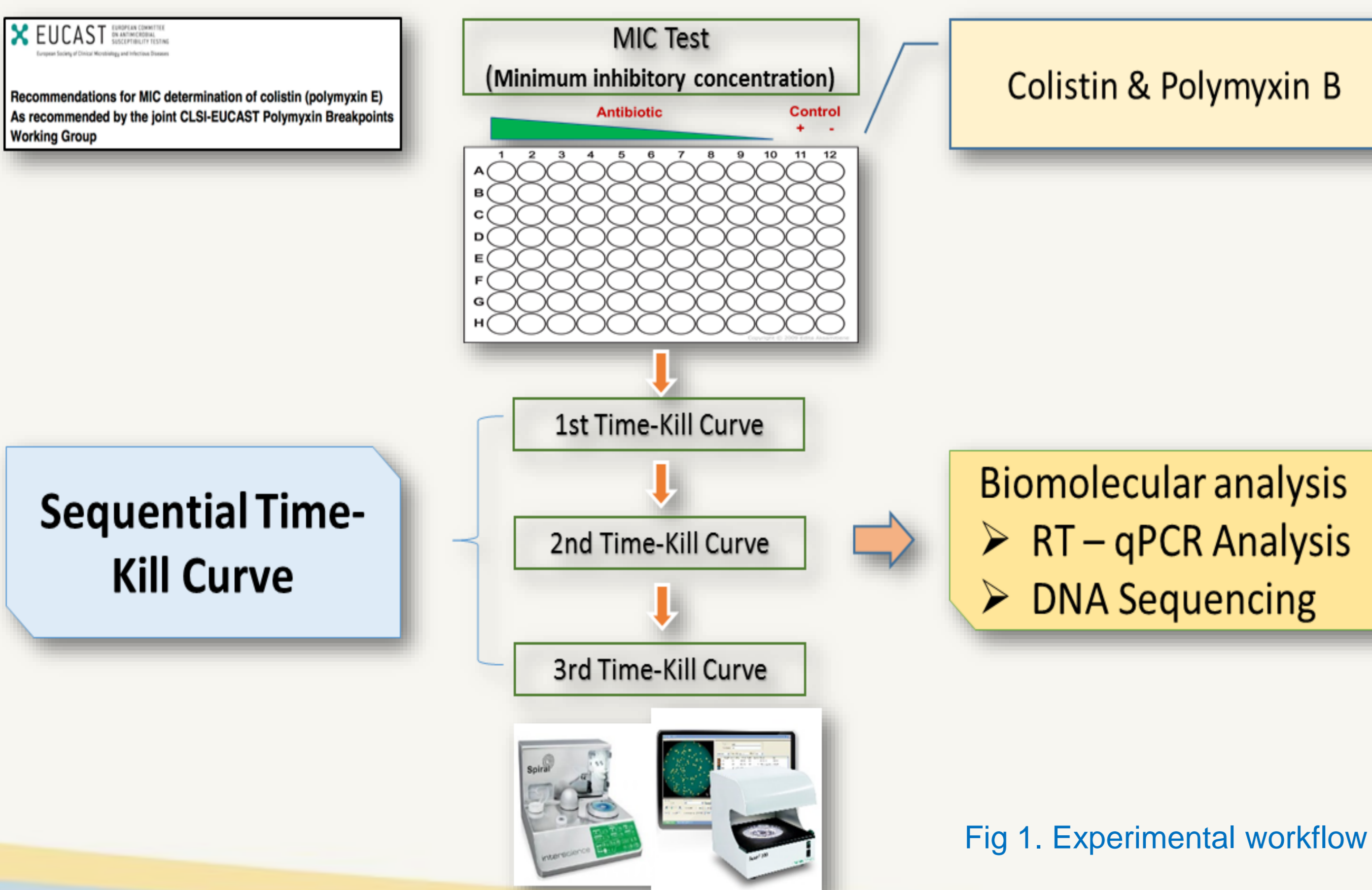


Fig 1. Experimental workflow

### Methods

- The inoculum used in 2<sup>nd</sup> or 3<sup>rd</sup> time-kill curve (TKC) were taken from the previous TKC obtained from the regrowth bacteria at maximal antibiotic concentration
- 1<sup>st</sup> Time-kill study :
  - Concentrations: 0,25; 0,5; 1; 2 and 4\*MIC
- 2<sup>nd</sup> and 3<sup>rd</sup> Time-kill study:
  - Concentrations: adapted based on the 1<sup>st</sup> and 2<sup>nd</sup> time-kill study, respectively.
- Analysis of heteroresistant subpopulations of bacteria by Population analysis profiles (PAPs) and MICs determination after TKC were performed as well

### Results

#### Sequential Time-Kill Curves\*

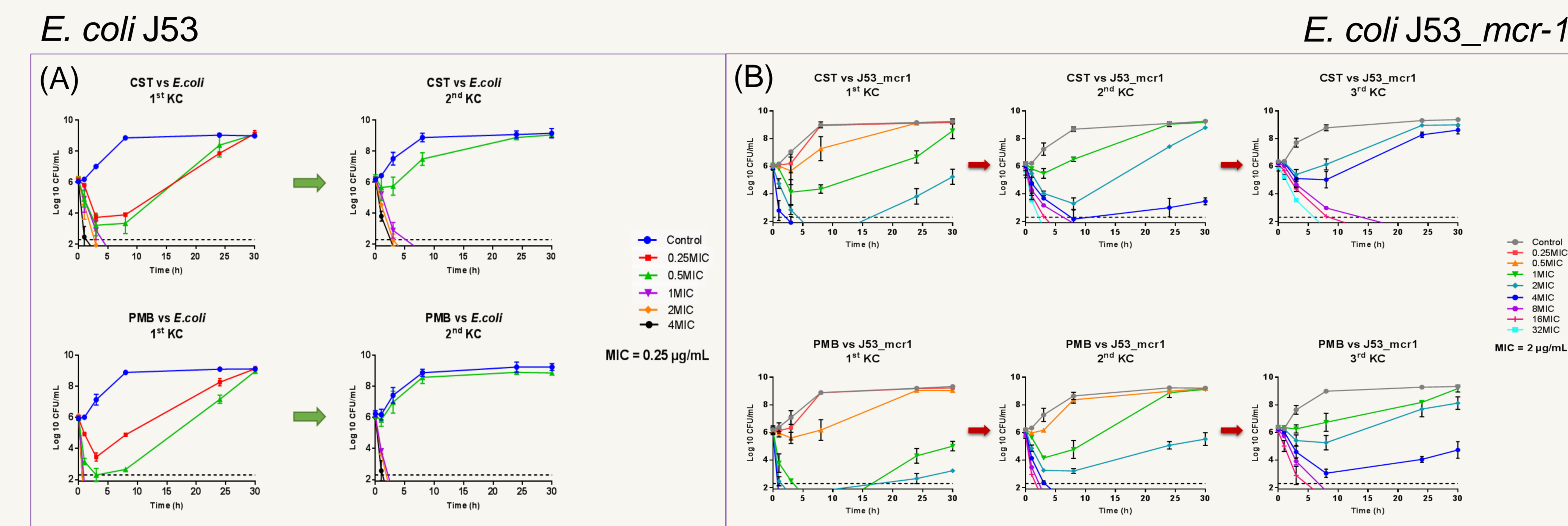


Fig 2. Time-Kill curves of colistin (CST) & polymyxin B (PMB) for *E. coli* J53 & *E. coli* J53 containing plasmid MCR-1 (J53\_ *mcr1*)

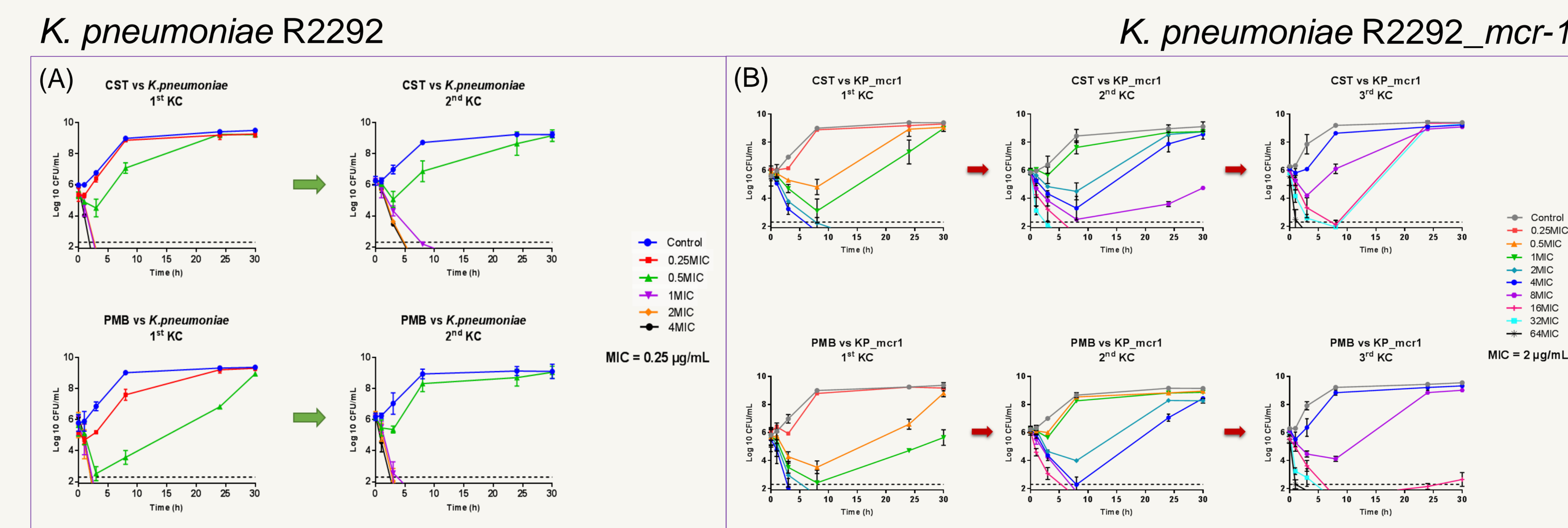


Fig 3. Time-Kill data of CST & PMB for *K. pneumoniae* & *K. pneumoniae* with *mcr-1* (KP\_ *mcr1*)

- Reference strains are not able to adapt the resistance to colistin & polymyxin B (only resistant subpopulation) (Fig. 2A & 3A)

\*Data are presented as means and standard deviations after two times replications

### Results

- The presence of MCR-1 leads to high-level polymyxins resistance (Fig. 2B, 3B & 4)

Strain	Colistin	Polymyxin B	
J53	0,25	0,125	
J53 - 1st TKC	0,25	0,25	
J53 - 2nd TKC	0,25	0,25	
J53_ <i>mcr1</i>	2	2	LOW Level Resistance
J53_ <i>mcr1</i> - 1st TKC	8	4	HIGH Level Resistance
J53_ <i>mcr1</i> - 2nd TKC	16	8	
J53_ <i>mcr1</i> - 3rd TKC	32	16	
KP	0,25	0,25	
KP - 1st KC	0,25	0,25	
KP - 2nd KC	0,25	0,25	
KP_ <i>mcr1</i>	2	2	LOW Level Resistance
KP_ <i>mcr1</i> - 1st TKC	16	8	HIGH Level Resistance
KP_ <i>mcr1</i> - 2nd TKC	64	16	
KP_ <i>mcr1</i> - 3rd TKC	512	128	

Fig 4. MICs (mg/L) confirmation after sequential Time-Kill Curve

- PAPs confirmed the absence of heteroresistant subpopulation since both antibiotics exhibited bacterial killing above 0.125 mg/L and 4 mg/L concentration for reference strain and their transconjugants respectively

### Conclusion

Reference strains did not show adaptive resistance to polymyxins while the presence of MCR-1 facilitated the step-by-step adaptation. These findings suggest that plasmid MCR-1 favor selection of another resistance mechanism, which may lead to develop high resistance to polymyxins. Biomolecular studies are running to determine mechanisms involved.

### References

- Expert Rev Anti Infect Ther. 2012
- Jayol, A *et al.*, Eurosurveillance 21. 2016
- Cheng, Y.-H. *et al.*, Antimicrob. Agents Chemother. 2015
- Liu Y.-Y. *et al.*, Lancet Infect Dis. 2016
- European Committee on Antimicrobial Susceptibility Testing. 2016